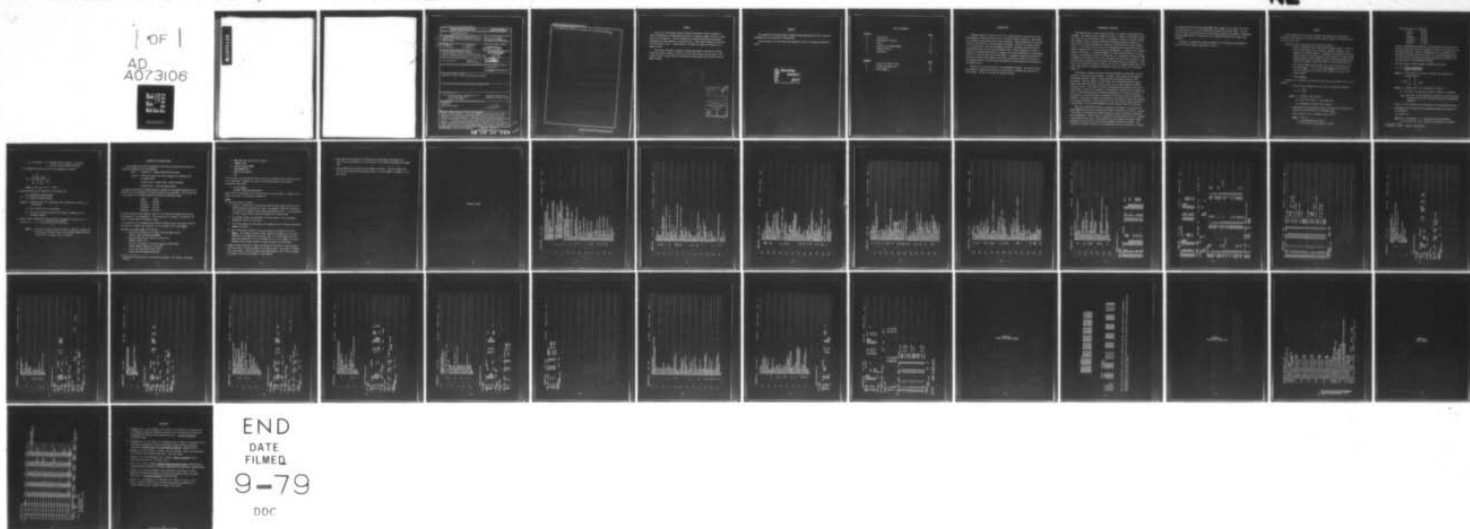


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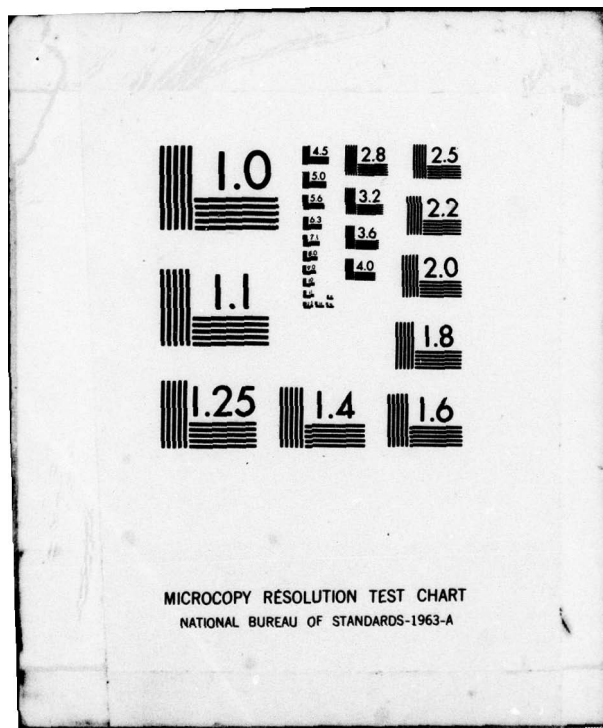
AEROSPACE MEDICAL RESEARCH LAB WRIGHT-PATTERSON AFB OH F/G 6/16
A COMPUTERIZED METHOD FOR DETERMINATION OF REGIONAL BLOOD FLOW --ETC(U)
MAR 79 K C SMITH, S WARD, K J GREENLEES F33615-76-C-5001
AMRL-TR-79-1 NL

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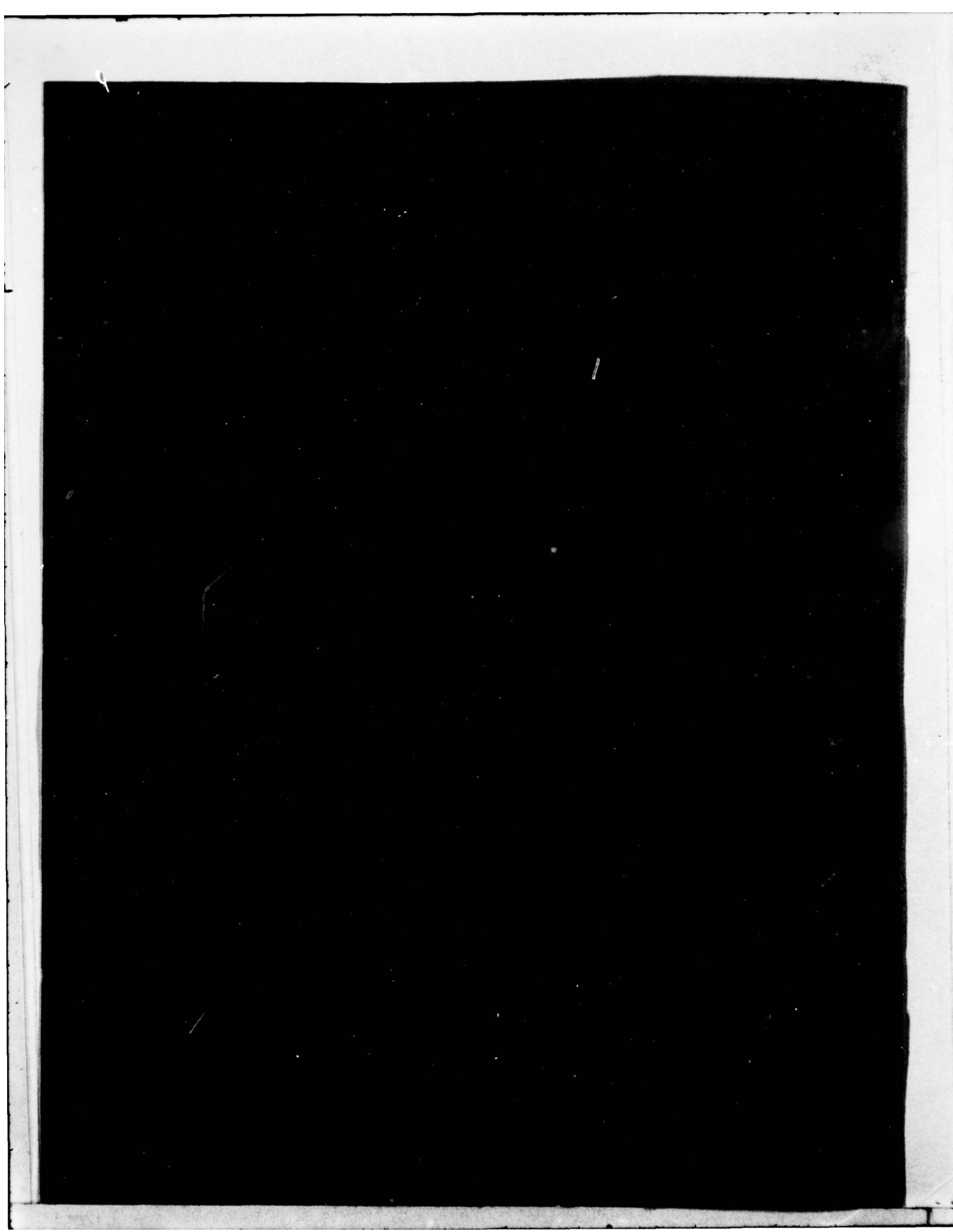
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Block 20. Abstract (cont'd)

7 program, execution of the program, and the program listing. 4

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SUMMARY

Studies in the Aerospace Medical Research Laboratory utilize radioactive tracer microspheres to determine blood flow to a specific region, tissue or organ under $+G_z$ acceleration. The microspheres are injected and become distributed throughout the body similar to red blood cells. They become trapped in capillary beds according to size. Tissue samples are taken and radiation measured in a gamma counter.

This report describes a computer program developed to calculate corrected counts per minute, regional blood flow, and cardiac output from raw data. The program offers a rapid, easily used method for processing data obtained from the gamma counter.

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PREFACE

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A *laboratory*
M *director's*
R *fund*
L

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INTRODUCTION

Studies similar to those of Sostre (7) utilizing plastic radioactive tracer microspheres to calculate blood flow to a specific region, tissue, or organ under $+G_z$ acceleration are being done in this Laboratory. The microspheres are injected into the left ventricle, and distributed throughout the body in a manner similar to red blood cells. They are entrapped in specific capillary beds according to blood distribution and the size microsphere used. Tissue samples from areas of interest are weighed and placed in a gamma counter for determination of microsphere density in terms of counts per minute (CPM). A computer program was developed to calculate corrected CPM, regional blood flow, and cardiac output from raw data. This program offers a rapid, easily used method for processing data obtained from the gamma counter.

Section 2 is a brief outline of the experimental design. The theory behind the program is presented in Section 3, and Section 4 contains the execution of the program. Section 5 includes the program listing.

EXPERIMENTAL PROCEDURE

The animals used, in this instance baboons, are lightly anesthetized with ketamine. Two femoral artery cutdowns are made; a catheter is inserted into the left ventricle via one femoral artery. Microsphere injections are made through this catheter using a remotely controlled microsphere injection system (2). A small silicone coated polyethylene catheter is inserted into the other femoral artery. It is placed with the tip in the descending aorta just below the arch. Fifteen seconds prior to the injection of microspheres, a constant rate reference sample is withdrawn. A Harvard Pump (R) (7) is used in this Laboratory, but any constant rate withdrawal system is acceptable. (6) Withdrawal of blood continues throughout the injection series until one minute after the final injection. The volume of blood withdrawn is calculated, and radioactivity is determined. Blood flow to this reference sample is taken to be that of a "known" organ; flow to any other organ then be calculated by relating to the blood flow of the reference sample (1).

The animal is placed in the Oloff Primate Restraint System (PRS) (5), and allowed to recover from the ketamine. The PRS is then placed on the Dynamic Environment Simulator (DES), a large man-rated centrifuge located in this Laboratory. When the animal is fully awake, the acceleration profile begins. Peak acceleration is either at $+3G_z$ or $+5G_z$, with an equal number of animals exposed to each. Five types of 15 micron radiolabeled microspheres are used for each animal: Cerium 141, Chromium 51, Strontium 85, Scandium 46, and Iodine 125. 0.025 to 0.050 microcuries are injected, depending on the relative activity of the isotope. The first isotope is injected at $+1G_z$ and is used to establish a baseline. When peak acceleration is reached, the remaining isotopes are injected at 30 second intervals. After injection of the last isotope, peak acceleration is maintained for 30 seconds bringing total $+G_z$ exposure time to 120 seconds.

When the acceleration profile is completed, the animal is sacrificed with Uthanol (R) and the brain is removed and fixed in formalin for 72 hours. One half of the brain is sectioned into various regions, such as brainstem, cerebellum, and hypothalamus. The other half is placed in several vials and used to estimate total cerebral blood flow. Tissue samples are also taken from various organs throughout the body, with each location assigned a specific ID number. All samples are weighed and placed in an appropriate vial to be counted in a Packard

Auto-Gamma Scintillation Spectrometer[®]. When a sample is too large to fit into one counting vial, it can be sectioned and placed into several vials. All the vials can then be incorporated into a group and treated as one sample. Information from the gamma counter is printed out on a standard teletype with a punch tape capacity.

Disposal of radioactive animal carcasses is in accordance with Appendix B, Title 10, Code of Federal Regulations, Part 20.

THEORY

Program GAMMA was written for a Control Data Corporation 6600 computer located at the Aeronautical Systems Division Computer Center at Wright-Patterson Air Force Base, Ohio.

There are three steps involved in using this program:

1. Create a permanent file of data from the gamma counter. This file will contain counts of all reference samples, standards, assay samples, and residue samples. Standards are aliquots of the microsphere solution of the same age as those injected into the animals. Residue samples include samples from the withdrawn catheters, post-experiment syringes, injection catheters, and manifold washouts.
2. Run program GAMMA using the above file, which is given the local file name Tape 1. A local file named Tape 2 will be created, this file contains the corrected CPM, regional blood flow, and cardiac output.
3. Catalog Tape 2.

Computation of corrected CPM, regional blood flow, and cardiac output is as follows:

1. Counts from all samples are corrected for background radiation:

$$c'_1 = c_1 - \frac{e_1^2}{c_1}$$

where c_1 = original count, window 1

e_1 = mean count window 1, from empty vials

c'_1 = background corrected count, window 1

2. Counts made on days previous to the assay counts are corrected for radioactive decay during the intervening time:

decay corrected count = original count $\times e^{(ct/h)}$

where $c = \ln(.5)$

t = days elapsed since count

h = half-life of given isotope, in days

Half-life values (4) used are:

Iodine	60 days
Cerium	32 days
Chromium	28 days
Strontium	64 days
Scandium	84 days

Counts of standards are corrected for decay using the same half-life for each window; the isotope is assumed to decay at the same rate over the entire spectrum. Counts of samples that are not standards are corrected for decay using the half-life of the corresponding isotope for each window; the decay rate within each window is assumed to be the decay rate of the predominant isotope.

3. The total amount of each isotope is computed as follows:

$$z_i = y_i \times \frac{\text{volume injected (ml)}}{\text{volume counted (ml)}}$$

where z_i = total amount of isotope i injected, not corrected for residue samples

$$y_i = \frac{1}{n_i} \sum_{k=1}^5 \sum_{j=1}^5 c_{ijk}$$

where y_i = average count of the standard for isotope i

n_i = the number of times the count for isotope i is repeated

c_{ijk} = the count of the standard for isotope i in the jth window repeated the kth time, corrected for decay and background radiation

Thus y_i is the average of the total counts in all windows from isotope i.

4. Corrected counts (corrected for overlapping windows) are computed by the formula*:

$$T = (F'F)^{-1} F'C$$

where T is a 5x1 matrix; t_i = corrected counts from isotope i

F is a 5x5 matrix; f_{ij} = proportion of isotope i in window j

* Rosenblatt, Judah. Personal communication

C is a 5x1 matrix; c_j = observed count in window j, corrected
for decay and background radiation

F is estimated from the counts of the standards as follows:

$$f_{ij} = \frac{\frac{1}{n_i} \sum_{k=1}^{n_i} c_{ijk}}{\sum_{i=1}^5 \frac{1}{n_i} \sum_{k=1}^{n_i} c_{ijk}}$$

where n_i and c_{ijk} are as in 3 above

5. Regional blood flow is computed by the formula (5):

$$g_i = \frac{t_i \times \text{withdrawal rate (ml/min)}}{r_i \times \text{weight of sample (grams)}}$$

where g_i = regional blood flow immediately after injection of isotope i, in
[ml/min/gram]

t_i = the corrected count from isotope i

r_i = the sum of corrected counts from isotope i, summed over all
reference samples

6. Cardiac output immediately after injection of isotope i is given by the

$$\text{formula (3): } s_i = \frac{w_i \times \text{withdrawal rate (ml/min)}}{r_i}$$

where w_i = the total corrected count of isotope i injected, computed by
subtracting corrected counts of all residue samples from the
total count of isotope i (z_i in 3 above).

EXECUTION OF PROGRAM GAMMA

The permanent data file described in the theory section should contain the following information for each sample:

Line 1 Columns 1-6: sample identification number

Lines 2-6 Counts for each of the five isotopes in ascending order by energy level

Columns 15-20: energy level, center of window

Columns 50-55: count for given window

The sample identification number should be unique; no two samples should have the same ID, with the exception of multiple counts of the standards. The energy level at the center of the window should be within the following ranges:

Iodine 29-35

Cerium 142-148

Chromium 317-323

Strontium 511-517

Scandium 1117-1123

The above format is the standard output of the Packard Auto-Gamma Scintillation Spectrometer R (see Appendix A). The program can be changed to accommodate the use of a counter with a different format.

If the data from an experiment are on several different paper tapes they should be read into the computer and combined into one permanent file using Editor.

Execution of program GAMMA is as follows:

1. Attach the permanent file of data from the gamma counter
ATTACH, TAPE1, PFN,CY=XX,ID=XXXXXX
2. Request permanent file space for output file
REQUEST, TAPE2, *PF
3. Attach the permanent file which contains program GAMMA
ATTACH, GAMMA, WARD35,CY=1,ID=L740530
4. Attach the subroutine library IMSL*
ATTACH,IMSL,ID=LIBRARY,SN=ASD,MR=1

* International Mathematical & Statistical Libraries. ASD Computer Subprogram Library

5. Make IMSL part of the local library

LIBRARY, IMSL

6. Compile program GAMMA

FTN(I=GAMMA,L=0)

7. Execute the program

LGO

At this point, the program will ask for certain information which should be typed in as requested. Appendix B contains an abbreviated sample with typed-in information underlined.

8. Catalog Tape 2

CATALOG, TAPE2, PFN, CY=XX, RP=999

where PFN is the permanent file name and XX is the cycle number. A sample of the output from Tape 2 is included in Appendix C.

NOTES:

1. Entering sample ID numbers

- a. A group of numbers may be entered by putting a minus sign in front of the selected number satisfying the range. For example, "580 -592" specifies all thirteen ID numbers between and including 580 and 592. It is important to leave a space between the first number and the minus sign.
- b. If too many numbers are specified, the extra numbers will be dropped starting at the end of the list.
- c. If too few numbers are specified, the program will not continue until more numbers are given.
- d. Do not include ID numbers in the list which do not exist in the file Tape 1. For example, suppose a group consists of samples 33 -45, and 47 and there is no sample 46 in the data file. If the ID numbers for the group are specified as "33 -47" instead of "33 -45 47" sample 47 will be dropped from the group because too many ID numbers are specified.

2. Samples counted previous to assay are grouped by the number of days elapsed from the count of the given sample to the assay counts. Any number of groups may be specified. After all groups have been entered, enter 0 for the number of samples, and 0 for the number of days elapsed.

3. When sample ID and weight are requested for each sample, the sample ID to enter is the identification of the location in the animal from which the sample came.
4. Assay samples may be grouped in any number of groups. After the sample IDs for the last group have been entered, enter 0 for the number of samples in the next group.

PROGRAM LISTING

```

1  PROGRAM GAMMA(INPUT,OUTPUT,TAPE1,TAPE2)
   DIMENSION BACK(5),CVOL(5),TVOL(5),Q(5,5)
   DIMENSION REF(5),NS(50),KCP(200),NGP(200),KGP(200,2)
   DIMENSION KSI(50),NTOT(5)
   DIMENSION NSDEL(200),DAY(200)
   DIMENSION MGPI(200)
   DIMENSION KOUN(200),TW(5),ICT(5),LHM(5),KCT(5),RCT(5)
   DIMENSION TOT(5),SSGP(200,5),FLOM(5),KREF(20),PER(5,5)

10  * TO CHANGE # ISOTOPE
   * CHANGE DATA STATEMENTS FOR LHM, REF AND BACK
   * CHANGE DIMENSIONS OF SSGP,PER AND Q
   * CHANGE VALUE OF "NIS" -- # OF ISOTOPE
   * INCREASE DIMENSIONS OF ALL RELEVANT VARS IF NECESSARY
   * CHANGE DIMENSIONS IN SUBROUTINES INV AND ICLS

15  *
   DATA LHM/32,145,320,514,1120/
   DATA LHM/32,320,514,1120,6/
   DATA REF/SHI125,SHCE145,SHCR141,SHSR65,SHSC46 /
   DATA REF/SHI125,SHCR141,SHSR65,SHSC46,SH /
   DATA BACK/79,74,19,32,4,7/
   PRINT 1
   FORMAT(*10CORRECTED MICROSPHERE COUNTS*/)
   NIS=4
   NIS=5
   PRINT 400,NIS,(REF(I),I=1,NIS)
   FORMAT(* NUMBER OF ISOTOPE ASSUMED TO BE*,I3/
1  * ISOTOPE *,10A6)
   PRINT 5
   FORMAT(* ENTER SUBJECT ID *)
   READ 3,SUBJ
   PRINT 2
   FORMAT(* ENTER EXPERIMENT DATE *)
   READ 3,DATE
   PRINT 4
   FORMAT(* ENTER ASSAY DATE *)
   READ 3,ASSAY
   FORMAT(A10)
   WRITE(2,107)SUBJ,DATE,ASSAY
   FORMAT(3A10)
   IT=0
   DO 62 I=1,NIS
   PRINT 7,I,REF(I)
   FORMAT(* ISOTOPE *,I2,1X,A5)
   PRINT 403
   FORMAT(5X,*ENTER VOLUME COUNTED *)
   READ *,CVOL(I)
   PRINT 404
   FORMAT(5X,*ENTER VOLUME INJECTED *)
   READ *,TVOL(I)
   PRINT 405
   FORMAT(5X,*ENTER # SAMPLE IDS *)
   READ*,NKS
   IF(NKS.LE.0)GO TO 62
   PRINT 14
   CALL NUHBNKS,ADUN)
   DO 63 J=1,NKS

```



```

17=IT+1
KST(I)=KOUN(J)
63 KST(I)=I
62 CONTINUE
120 PRINT 120
FORMAT(* ENTER # REFERENCE SAMPLES *)
CALL REFIO(REF,KREF,RCF,NIS)
65 PRINT 6
FORMAT(* ENTER WITHDRANAL RATE, REFERENCE SAMPLE *)
READ *,WITHD
PRINT 10
10 FORMAT(* ENTER # SAMPLES FROM RESIDUE *)
READ *,NCATH
IF(NCATH.LE.0)GO TO 110
PRINT 14
CALL NUMB(NCATH,KC)
110 PRINT 108
108 FORMAT(* ENTER # SAMPLES COUNTED PREVIOUS TO ASSAY*)
I=0
53 PRINT 50
FORMAT(IX,* ENTER # SAMPLES AND # DAYS ELAPSED *)
50 READ *,NSS,DDD
IF(NSS.LE.0)GO TO 51
PRINT 14
CALL NUMB(NSS,KOUN)
DO 52 K=1,NSS
I=I+1
NSDEL(I)=KOUN(K)
52 DAY(I)=DDD
GO TO 53
51 NOEL=I
NGPS=IE=0
IB=I=1
16 PRINT 11,I
11 FORMAT(* ENTER # SAMPLES IN GROUP*,13,1X)
READ *,NGP(I)
IF(NGP(I).LE.0)GO TO 13
NGPS=I
PRINT 14
FORMAT(IX,*ENTER SAMPLE IDS *)
CALL NUMB(NGP(I),KOUN)
IE=IB-1+NGP(I)
100 K=0
DO 15 J=IB,IE
K=K+1
KGP(J,1)=KOUN(K)
15 KGP(J,2)=I
IB=IE+1
I=I+1
GO TO 16
*
*
110 PROCESS STANDARDS AND REFERENCE SAMPLES
REWIND 1
MREF=0
DO 130 I=1,NTS
NTOT(I)=0

```

```

115      TOT(I)=0.
      DO 130 J=1,NIS
130      PER(I,J)=0.
135      READ(I,17)IID,((IM(I),ICT(I)),I=1,NIS)
17      FORMAT(I6/(14X,I6,29X,I6))
      IF(EOF(I))131,132
      CHECK IF A STANDARD
      DO 133 I=1,IY
132      IF(IID.NE.MS(I))GO TO 133
      CALL WINDOW(IID,IN,ICT,LIMM,NIS,BACK)
      IS=MS(I)
      IF(IS.LE.0.OR.IS.GT.NIS)STOP "INDEX"
      CALL HALF(IID,ICT,MODEL,NSDEL,DAY,IS,NIS)
      NTOT(IS)=NTOT(IS)+1
      DO 134 J=1,NIS
130      *
      TOT(IS) = SUM FOR ISOTOPE IS OVER ALL WINDOWS
      TOT(IS)=TOT(IS)+ICT(J)
      PER(IS,J)=PER(IS,J)+ICT(J)
      GO TO 135
135      *
      CONTINUE
      CHECK IF A REFERENCE
      IF(REF.LE.0)GO TO 135
      DO 136 I=1,NREF
140      IF(IID.NE.REF(I))GO TO 136
      CALL HALF(IID,ICT,MODEL,NSDEL,DAY,0,NIS)
      CALL WINDOW(IID,IN,ICT,LIMM,NIS,BACK)
      NREF=NREF+1
      DO 137 J=1,NIS
145      RCT(J)=RCT(J)+ICT(J)
137      GO TO 135
136      CONTINUE
      GO TO 135
131      DO 150 I=1,NIS
150      SUM=0.
      DO 151 J=1,NIS
151      SUM=SUM+PER(I,J)
      IF(SUM.GT.0.160 TO 150
      PRINT 309,I
155      FORMAT(" NO STANDARD FOUND FOR ISOTOPE",I3/
      1 * ENTER CPN FOR EACH WINDOW")
      DO 301 J=1,NIS
      PRINT 302,J
160      FORMAT(5X,"WINDOW ",I2,1X)
161      READ *,PER(I,J)
      TOT(I)=TOT(I)+PER(I,J)
      NTOT(I)=NTOT(I)+1
      PER(I,J) IS THE COUNT OF ISOTOPE I IN WINDOW J
165      CONTINUE
      *
      COMPUTE FREQUENCY ARRAY
      DO 65 I=1,NIS
      IF(NTOT(I).LE.0)STOP "STANDARD"
      XX=1./NTOT(I)
      TOT(I)=TOT(I)*XX
170

```



```

DO 65 J=1,NIS
PER(I,J)=PER(I,J)*XX
IF(NREF.EQ.NREF)GO TO 160
PRINT 161,NREF,NREF
FORMAT(0 REF SAMPLES FOUND*,I3,5X,0 SPECIFIED*,I3)
CALL REFID(NREF,KREF,RCF,REF,NIS)
GO TO 13
175
180 DO 140 I=1,NIS
SUN=0
DO 141 J=1,NIS
SUN=SUN+PER(I,J)
SUN=1./SUN
DO 142 J=1,NIS
PER(I,J)=PER(I,J)*SUN
CONTINUE
CALL INV(REF,NIS,0)
COMPUTE CORRECTED CPHS FOR REFERENCE SAMPLES
190 DO 143 I=1,NIS
ICT(I)=RCF(I)
CALL CPM(CT,CT,PER,NIS,0)
PRINT 61,ICT(I),I=1,NIS)
PRINT 402,(RCF(I),I=1,NIS)
FORMAT(/% REF CPM %5I10)
411 FORMAT(% CORR CPM %5F10.0)
CO 411 I=1,NIS
TOT(I)=TOT(I)+VOLI(I)/CVOL(I)
PRINT 500,TOT
511 FORMAT(/% SYRINGE*,5612.5/)
PROCESS ASSAY SAMPLES AND RESIDUE SAMPLES
180
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230      31 J=1,NIS
      TOT(J)=TOT(J)-XCT(J)
      GO TO 25
      30 CONTINUE
      34 CHECK IF IN A GROUP
      IF(IE.LE.0)GO TO 35
      235 DO 33 I=1,IE
      IF(IID.NE.KGP(I,1))GO TO 33
      IGP=KGP(I,2)
      PRINT 230,IID,IGP
      240 POFMAT(I5,* IN GROUP*,I3)
      DO 36 J=1,NIS
      SSGP(IGP,J)=SSGP(IGP,J)+ICT(J)
      36 MGP(IGP)=MGP(IGP)+1
      GO TO 25
      33 CONTINUE
      245 PRINT 40,IID
      POFMAT(* SAMPLE*,I5,* -- ENTER SAMPLE ID AND WEIGHT (G) *)
      40 READ *,IID,WT
      DO 42 I=1,NIS
      FLOW(I)=0.
      250 IF(IGT(I).EQ.0.0)GO TO 42
      FLOW(I)=XCT(I)*WTHD/(KGT(I)*WT)
      CONTINUE
      42 WRITE(2,44)LET,IID,KID,WT,(XCT(I),I=1,NIS)
      44 POFMAT(A1,2I5,6G12.5)
      255 WRITE(2,401)(FLOW(I),I=1,NIS)
      401 POFMAT(23X,5G12.5)
      GO TO 25
      180 LET=ING
      IFL=0
      260 IF(IE.LE.0)GO TO 333
      IF(IE.LE.0)GO TO 333
      DO 200 I=1,NGPS
      IF(MGP(I).EQ.0)GO TO 203
      IFL=1
      265 PRINT 201,I,MGP(I),NGP(I)
      POFMAT(* GROUP*,I3,* # SAMPLES FOUND*,I3,* # SPECIFIED*,I3)
      203 IF(IFL.EQ.0)GO TO 204
      PRINT 11,I
      READ *,NGP(I)
      PRINT 14
      CALL NUMB(NGP(I),KOUN)
      270 IE=IE-1+NGP(I)
      K=0
      DO 202 J=IE,IE
      K=K+1
      275 KGP(J,1)=KOUN(K)
      KGP(J,2)=I
      202 IE=IE+1
      GO TO 200
      280 IE=IE+NGP(I)
      280 CONTINUE
      IF(IFL.NE.0)GO TO 180
      285 DO 198 I=1,NGPS
      DO 191 J=1,NIS
      191 IGT(J)=SSGP(I,J)

```



```

CALL CPM(ICT,XCT,PER,NIS,Q)
PRINT 192,1
FORMAT(' GROUP #,I3,' -- ENTER SAMPLE ID AND WEIGHT (G) *')
READ *,XID,WT
DO 193 J=1,NIS
  FLOW(J)=0.
  FLOW(J)=XCT(J)-WTND/(RCT(J)*WT)
CONTINUE
WRITE(2,44)LET,I,XID,WT,XCT(J),J=1,NIS)
WRITE(2,401)(FLOW(J),J=1,NIS)
CONTINUE
CARDIAC OUTPUT
PRINT 240,(TOT(J),J=1,NIS)
FORMAT(' CPM OF ISOTOPE INJECTED*/5X,5612.5)
DO 241 I=1,NIS
  IF (RCT(I)-EQ.0.0)GO TO 220
  TOT(I)=TOT(I)+WTND/RCT(I)
GO TO 241
220 TOT(I)=0.
241 CONTINUE
PRINT 242,(TOT(J),J=1,NIS)
FORMAT(' CARDIAC OUTPUT AT EACH ISOTOPE (HL/MIN)*5X,5612.5)
LET=1MS
WRITE(2,44)LET
LET=1HF
WRITE(2,243)LET,SUBJ,DATE,ASSAY,(TOT(J),J=1,NIS)
FORMAT(1,X,3A10,5612.5)
CALL EXIT
END

```

SYMBOLIC REFERENCE MAP (R=1)

ENTRY POINTS
10266 GAMMA

VARIABLES	SN	TYPE	RELOCATION	12332	BACK	REAL	ARRAY
12302 ASSAY		REAL		12301	DATE	REAL	
12337 CYOL		REAL	ARRAY	12332	DOO	REAL	
14930 DAY		REAL	ARRAY	12277	I	INTEGER	
17566 FLOW		REAL		15665	ICT	INTEGER	ARRAY
12317 IG		INTEGER		12331	IFL	INTEGER	
12316 IG		INTEGER		12321	IID	INTEGER	
12326 IGP		INTEGER		12303	IT	INTEGER	
12322 IS		INTEGER		12305	J	INTEGER	
19560 IW		INTEGER	ARRAY	12471	KC	INTEGER	ARRAY
12313 K		INTEGER		13311	KGP	INTEGER	ARRAY
15350 KDUH		INTEGER	ARRAY	17673	KREF	INTEGER	ARRAY
12327 KID		INTEGER		14131	KST	INTEGER	ARRAY
12407 KS		INTEGER	AFRAY	15672	LIM	INTEGER	ARRAY
12325 LET		INTEGER		12320	MREF	INTEGER	
15049 MGP		INTEGER	ARRAY	12314	NOEL	INTEGER	
12310 NCATH		INTEGER					

LOOPS	LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES
10505	130	I	113 117	158	NOT INNER
10514	130	J	116 117	28	INSTACK
10525	133	I	118 118	108	EXT REFS
10542	133	I	122 136	338	EXT REFS
10564	134	J	129 134	58	EXT REFS
10577	136	I	139 147	229	EXT REFS
10611	137	J	144 145	38	EXT REFS
10622	150	I	149 164	418	EXT REFS
10630	151	J	151 152	38	INSTACK
10640	331	I	157 161	168	EXT REFS
10684	65	I	168 173	248	EXT REFS
10701	65	J	172 173	38	INSTACK
10717	140	I	179 186	308	NOT INNER
10725	141	J	181 182	38	INSTACK
10740	142	J	184 185	38	INSTACK
10753	143	I	189 190	38	INSTACK
10775	411	I	196 197	38	INSTACK
11010	22	I	205 208	148	NOT INNER
11016	22	I	207 208	28	INSTACK
11030	23	I	210 210	108	EXT REFS
11046	23	I	215 217	58	INSTACK
11056	171	I	220 222	58	EXT REFS
11072	30	I	227 232	158	EXT REFS
11100	31	J	229 230	38	INSTACK
11112	33	I	235 244	258	EXT REFS
11125	36	J	240 241	48	INSTACK
11146	42	I	248 252	58	INSTACK
11174	206	I	262 281	538	EXT REFS
11232	202	J	274 277	48	EXT REFS
11251	190	I	283 297	458	EXT REFS
11256	191	J	284 285	38	INSTACK
11273	193	J	290 294	58	INSTACK
11326	241	I	301 306	58	INSTACK

STATISTICS

PROGRAM LENGTH 74748 3900
 BUFFER LENGTH 102608 4272
 520008 CM USED

```

1  SUBROUTINE CPM(ICT,XCT,PER,NIS,Q)
2  DIMENSION ICT(NIS),XCT(NIS),PER(NIS,NIS)
3  DIMENSION Q(NIS,NIS)
4  PER IS INV(A *A TRANS) * A
5  WHERE A(I,J) IS THE PROPORTION OF COUNTS FROM ISOTOPE
6  I IN WINDOW J
7  SUM OF A(I,J) OVER J FOR FIXED I IS 1.0
8  DO 5 I=1,MIS
9  DO 4 J=1,NIS
10  XCT(I)=XCT(I)*PER(I,J)*ICT(J)
11  CONTINUE
12  CALL ICLS(Q,XCT,NIS)
13  RETURN
14  END

```

SYMBOLIC REFERENCE MAP (R=1)

ENTRY POINTS
3 CPM

22

VARIABLES	SN	TYPE
54 I		INTEGER
55 J		INTEGER
0 PER		REAL
0 XCT		REAL

RELOCATION	ARRAY	F.P.
	ARRAY	F.P.

0	ICT	INTEGER	ARRAY	F.P.
0	NIS	INTEGER		F.P.
0	Q	REAL	ARRAY	F.P.

EXTERNALS	TYPE	ARGS
ICLS		3

STATEMENT LABELS

50

LOOPS	LABEL	INDEX
20	5	I
30	4	J

FROM-TO	LENGTH	PROPERTIES	NOT INNER
0 12	178		
10 11	48	INSTACK	

STATISTICS

PROGRAM	LENGTH	520008	CM	USED
141517G5				

728 58


```

1  SUBROUTINE NUMB(NLIN,KC)
   DIMENSION KC(200)
   I=0
   15 READ *,KK
   IF(KK.GT.0)GO TO 11
   IF(I.LE.0)GO TO 12
   IB=KC(I)+1
   IE=IABS(KK)
   DO 13 J=IB,IE
     I=I+1
   13 KC(I)=J
     GO TO 14
   12 KK=IABS(KK)
     I=I+1
   11 KC(I)=KK
     IF(I-NLIN)15,20,21
   14 RETURN
   20 PRINT 22,I
   21 FORMAT(' SPECIFIED # IS ',I5)
   22 RETURN
   END

```

20

SYMBOLIC REFERENCE MAP (R=1)

ENTRY POINTS

3 NUMB

VARIABLES	SN	TYPE	RELOCATION
61 I	63	INTEGER	28
64 IE	65	INTEGER	J
0 KC	62	INTEGER	KK
8 NLIN		ARRAY	F.P.
			F.P.

FILE NAMES	MODE	INPUT	OUTPUT	FMT
	FREE			

INLINE FUNCTIONS	TYPE	ARGS
IABS	INTEGER	1
	INTRIN	

STATEMENT LABELS

STATEMENT LABELS	30	12	0	13
32 11				
36 14				
42 21				
	55	22		
			0	20
				INACTIVE

LOOPS LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES
24 13	J	9 11	28	INSTACK

STATISTICS

PROGRAM LENGTH	CM USED
52008	668 54

```

1      SUBROUTINE REFID(NREF,KREF,RCF,REF,NIS)
      DIMENSION KREF(1200),RCF(NIS),REF(NIS)
      DO 1 I=1,NIS
      RCF(I)=0.
      READ *,NREF
      PRINT 121
      121  FORMAT(' ENTER SAMPLE IDS FOR REFERENCE ')
      CALL NUNB(NREF,KREF)
      RETURN
      END
10

```

SYMBOLIC REFERENCE MAP (R=1)

ENTRY POINTS
3 REFID

VARIABLES	SN	TYPE	RELOCATION	0	KREF	INTEGER	ARRAY	F.P.
43 I	1	INTEGER		0	KREF	INTEGER		F.P.
8 NIS		INTEGER		0	NREF	INTEGER		F.P.
8 RCF		REAL		0	REF	REAL	ARRAY	F.P.

FILE NAMES INPUT MODE OUTPUT FMT

EXTERNALS NUMB TYPE ARGS

STATEMENT LABELS

LOOPS LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES
11 1	1	3 4	28	INSTACK

STATISTICS
PROGRAM LENGTH 528008 CH USED 448 36


```

1 SUBROUTINE WINDOW (IZO,IN,ICT,LIMW,NIS,BACK)
  DIMENSION BACKIN(5)
  DIMENSION IN(INIS),ICT(NIS),LIMW(NIS)
  DO 10 I=1,NIS
21 IF(IABS(IN(I)-LIMW(I)).LE.3)GO TO 10
  PRINT 19,IZO,IN(I),LIMW(I),ICT(I)
19 FORMAT(1X,WINCH WRONG/
  1 5X,SAMPLE,I3,0 WINCH,I2,0 CENTROID,I5/
  2 5X,SHOULD BE,I5,0 COUNT,I10)
  PRINT 20
20 FORMAT(0 ENTER CORRECT CENTROID AND COUNT *)
  READ *,IN(I),ICT(I)
  GO TO 21
  CONTINUE
  CORRECT FOR BACKGROUND COUNTS
  CO 1 I=1,NIS
  IF(ICT(I).LE.0)GO TO 1
  XX=BACK(I)*BACK(I)/ICT(I)
  XX=ICT(I)-XX
  IF(XX.LE.0)XX=0.
  ICT(I)=XX+.5
  CONTINUE
1 RETURN
ENDC

```

SYNOCLIC REFERENCE MAP (R=1)

ENTRY POINTS
3 WINDCM

VARIABLES		SN	TYPE	RELOCATION			
0	BACK		REAL	ARRAY	F.P.	126	INTEGER
0	ICT		INTEGER	ARRAY	F.P.	0	INTEGER
0	IN		INTEGER	ARRAY	F.P.	0	INTEGER
0	IN		INTEGER	ARRAY	F.P.	127	REAL

FILE NAMES	MODE	FREE	OUTPUT	FMT
1	1	1	1	1
2	2	2	2	2
3	3	3	3	3
4	4	4	4	4
5	5	5	5	5
6	6	6	6	6
7	7	7	7	7
8	8	8	8	8
9	9	9	9	9
10	10	10	10	10
11	11	11	11	11
12	12	12	12	12
13	13	13	13	13
14	14	14	14	14
15	15	15	15	15
16	16	16	16	16
17	17	17	17	17
18	18	18	18	18
19	19	19	19	19
20	20	20	20	20
21	21	21	21	21
22	22	22	22	22
23	23	23	23	23
24	24	24	24	24
25	25	25	25	25
26	26	26	26	26
27	27	27	27	27
28	28	28	28	28
29	29	29	29	29
30	30	30	30	30
31	31	31	31	31
32	32	32	32	32
33	33	33	33	33
34	34	34	34	34
35	35	35	35	35
36	36	36	36	36
37	37	37	37	37
38	38	38	38	38
39	39	39	39	39
40	40	40	40	40
41	41	41	41	41
42	42	42	42	42
43	43	43	43	43
44	44	44	44	44
45	45	45	45	45
46	46	46	46	46
47	47	47	47	47
48	48	48	48	48
49	49	49	49	49
50	50	50	50	50
51	51	51	51	51
52	52	52	52	52
53	53	53	53	53
54	54	54	54	54
55	55	55	55	55
56	56	56	56	56
57	57	57	57	57
58	58	58	58	58
59	59	59	59	59
60	60	60	60	60
61	61	61	61	61
62	62	62	62	62
63	63	63	63	63
64	64	64	64	64
65	65	65	65	65
66	66	66	66	66
67	67	67	67	67
68	68	68	68	68
69	69	69	69	69
70	70	70	70	70
71	71	71	71	71
72	72	72	72	72
73	73	73	73	73
74	74	74	74	74
75	75	75	75	75
76	76	76	76	76
77	77	77	77	77
78	78	78	78	78
79	79	79	79	79
80	80	80	80	80
81	81	81	81	81
82	82	82	82	82
83	83	83	83	83
84	84	84	84	84
85	85	85	85	85
86	86	86	86	86
87	87	87	87	87</

INLINE	FUNCTIONS	TYPE	ARGS
IABS		INTEGER	1
			INTRIN

STATEMENT LABELS					
	PMT	PMT	FMT	FMT	FMT
61 1	---	41 18	75 19	---	---
113 20	---	16 21	---	---	---

LCOPS	LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES	EXT	REFS
16	18	*	4 14	268			
51	1	1	16 22	118	OPT		

STATISTICS
PROGRAM LENGTH 5200B CM USED 150B 104


```

1 SUBROUTINE INV (PER,NIS,Q)
  DIMENSION PER(NIS,NIS),Q(NIS,NIS)
  DIMENSION A(5,5),M(10),B(5,5)

5 * COMPUTE INVERSE (PER * PER TRANS ) * PER
  *
  * A = PER * PER TRANS
  DO 1 I=1,NIS
    DO 1 J=1,NIS
      A(I,J)=0.
10 *
      DO 1 K=1,NIS
        A(I,J)=A(I,J)+PER(I,K)*PER(J,K)
      INVERSE A
15 *
      CALL LINV3F(A,B,1,NIS,NIS,D1,D2,M,IER)
      DO 4 I=1,NIS
        DO 4 J=1,NIS
          Q(I,J)=A(I,J)
4 *
      DO 2 I=1,NIS
        DO 2 J=1,NIS
          B(I,J)=0.
20 *
          DO 2 K=1,NIS
            B(I,J)=B(I,J)+A(I,K)*PER(K,J)
          DO 3 I=1,NIS
            DO 3 J=1,NIS
              PER(I,J)=B(I,J)
25 *
              RETURN
            END
  END

```

27

SYMBOLIC REFERENCE MAP (R=1)

ENTRY POINTS
3 INV

VARIABLES	SN	TYPE	RELOCATION	ARRAY	224 B	157 D2	160 IER	155 K	0 PER	212 W	F.P.	F.P.	ARRAY	ARRAY
161 A		REAL												
156 Q1		REAL												
153 I		INTEGER												
154 J		INTEGER												
0 NIS		INTEGER												
0 Q		REAL												

EXTERNALS
LINV3F

STATEMENT LABELS

LOOPS	LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES	NOT INNER	NOT INNER	INSTACK	NOT INNER
20	1	I	8 12	258					
21	1	J	9 12	228					
34	1	K	11 12	38					
52	4	I	15 17	158					

OPT=1

74/74

SUBROUTINE INV

LOOPS	LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES
61	4	J	16 17	38	INSTACK
70	2	* I	18 22	308	NOT INNER
71	2	* J	19 22	248	NOT INNER
106	2	K	21 22	48	INSTACK
121	3	* I	23 25	158	NOT INNER
130	3	J	24 25	38	INSTACK

STATISTICS

PROGRAM LENGTH 2768 190
52008 CH USED


```
1 SUBROUTINE ICLS(Q,BETA,N)
2   DIMENSION Q(N,N),BETA(N),Y(5),Z(5),B(5,5),M(5),ZND(5)
3   DIMENSION BINV(5,5),MK(10),C(5),SQ(5)
4   DO 2 I=1,N
5     DO 2 J=1,N
6       B(I,J)=0.
7     DO 1 I=1,N
8       Y(I)=0.
9       B(I,I)=1.
10      Z(I)=M(I)-BETA(I)
11      INO(I)=1
12      DO 4 I=1,N
13        DO 4 J=1,N
14          BINV(I,J)=B(I,J)
15          ICS=ILET=0
16          DO 30 I=1,N
17            M(I)=0.
18            DO 5 J=1,N
19              M(I)=M(I)+BINV(I,J)*Z(J)
20              IF (ABS(M(I)).LE.1.0E-8) M(I)=0.0
21              CONTINUE
22              IT=0
23              XMAX=0.
24              DO 6 I=1,N
25                IF (XND(I).GT.0) GO TO 6
26                IF (M(I).EQ.0.0) GO TO 6
27                ICS=1
28                IF (XMAX.GE.ABS(M(I))) GO TO 6
29                XMAX=ABS(M(I))
30                IT=I
31              CONTINUE
32              IF (ICS.GT.0) GO TO 7
33              DO 8 I=1,N
34                IF (M(I).GE.0.0) GO TO 8
35                ICS=2
36                IF (ABS(M(I)).LE.XMAX) GO TO 8
37                XMAX=ABS(M(I))
38                IT=I
39              CONTINUE
40              IF (ICS.LE.0) GO TO 100
41              DO 50 I=1,N
42                SQ(I)=0.
43              DO 9 J=1,N
44                SQ(I)=SQ(I)+Q(I,J)*BINV(I,J)
45                IF (ABS(SQ(I)).LE.1.0E-8) SQ(I)=0.
46              CONTINUE
47              DO 10 I=1,N
48                IF (SQ(I).NE.0.0) GO TO 11
49              CONTINUE
50              THET0=SIGN(1.0,M(IT))
51              GO TO 17
52              THET0=0.
53              DO 13 I=1,N
54                THET6=THET0+SQ(I)*BINV(I,I)
55                IF (ABS(THET0).LE.1.0E-8) GO TO 61
56                THET0=M(IT)/THET0
57                ITT=0
```

```

60      T0=1.0E20
        DO 14 I=1,N
          DEN=THET0*BINV(I,I)
          IF(CEN.LE.1.0E-8)GO TO 14
          IF(Y(I).EQ.0.0)GO TO 14
          DEN=Y(I)/DEN
          IF(DEN.GE.T0)GO TO 14
          ITT=I
          T0=DEN
14      CONTINUE
          IF(Y0.LE.1.0)GO TO 15
          THET=THET0
          ILET=1
          DO 21 I=1,N
            B(I,ITT)=SQ(I)
            GO TO 22
15      THET=THET0*T0
            ILET=2
            DO 23 I=1,N
              B(I,ITT)=0.
              9(I,ITT)=1.0
21      Y(I)=V(I)-THET*BINV(ITT,I)
            DO 25 I=1,N
              Z(I)=Z(I)-THET*SQ(I)
              IF(ABS(Z(I)).LE.1.0E-8)Z(I)=0.0
              IF(ABS(V(I)).LE.1.0E-8)V(I)=0.0
25      CONTINUE
            DO 24 I=1,N
              CO 24 J=1,N
                BINV(I,J)=B(I,J)
                CALL LINV3F(BINV,C,1,N,N,D1,D2,MK,IER)
                IF(ICS.EQ.1.AND.ILET.EQ.2)IND(ITT)=1
                IF(ICS.EQ.2.AND.ILET.EQ.1)IND(ITT)=0
                GO TO 3
100     CO 101 I=1,N
            DO 200 J=1,N
              BETA(I)=BETA(I)+Q(I,J)*Y(J)
              IF(ABS(BETA(I)).LE.1.0E-8)BETA(I)=0.
95      CONTINUE
          RETURN
        END

```

SYMBOLIC REFERENCE MAP (R=1)

ENTRY POINTS
3 ICLS

VARIABLES	SN	TYPE	RELOCATION	0	BETA	REAL	ARRAY	F.P.
457 B		REAL	ARRAY				ARRAY	
522 BINV		REAL	ARRAY	565 C		REAL		
440 DEN		REAL		442 D1		REAL		
443 D2		REAL		427 I		INTEGER		
431 ICS		INTEGER		444 IER		INTEGER		

VARIABLES	SN	TYPE	RELOCATION
432 ILET		INTEGER	
433 IT		INTEGER	
439 J		INTEGER	
441 THET		REAL	
437 T0		REAL	
553 WK		REAL	
445 Y		REAL	

EXTERNALS	TYPE	ARGS
515 IND	INTEGER	
436 ITT	INTEGER	
572 SQ	REAL	
435 THETO	REAL	
510 H	REAL	
434 XMAX	REAL	
452 Z	REAL	

INLINE FUNCTIONS	TYPE	ARGS	SIGN	REAL	2	INTRIN
ABS	REAL	1				

STATEMENT LABELS	ABS	REAL	2	INTRIN
0 1	0	2	65 .3	
0 4	0	5	127 6	
145 7	144 8		0 9	
0 18	207 11		0 13	
245 14	265 15		226 17	
0 21	304 22		0 23	
0 24	0 25		0 30	
0 28	283 61		365 100	
0 181	0 208			

31

LOOPS	LABEL	INDEX	FROM-TO	LENGTH	PROPERTIES
28 2	1	4 6	138		NOT INNER
25 2	1	5 6	28		INSTACK
63 1	1	7 11	68		INSTACK
51 4	1	12 14	148		NOT INNER
56 4	1	13 14	38		INSTACK
78 30	1	16 21	238		NOT INNER
77 5	1	18 19	48		INSTACK
128 6	1	24 31	109		OPT
136 8	1	33 39	78		INSTACK
151 68	1	41 46	258		NOT INNER
162 9	1	43 44	48		INSTACK
177 18	1	47 49	48		INSTACK
215 13	1	53 54	48		INSTACK
235 14	1	59 67	128		OPT
261 21	1	71 72	28		INSTACK
276 23	1	76 77	28		INSTACK
313 25	1	79 84	148		OPT
331 24	1	85 87	148		NOT INNER
336 24	1	86 87	38		INSTACK
366 181	1	92 96	228		NOT INNER
374 288	1	93 94	48		INSTACK

STATISTICS	PROGRAM	LENGTH	52008	ON	USED
		6238	403		

***** ZA4KGE4 *****
 ***** ZA4KGE4 *****
 ***** END OF LIST *****
 ***** END OF LIST *****

APPENDIX A OUTPUT FROM GAMMA COUNTER


```

000274 00001.0
000016 000160 000032 001662 000048 000108 000000 013791
000128 000086 000144 000221 000160 000212 000000 003312
000304 000096 000320 000216 000336 000243 000000 003388
000496 000048 000514 000050 000530 000048 000000 000790
001104 000041 001120 000045 001136 000050 000000 000750

```

	Counter ID	Counting time(min)	Centroid	Total CPM
Iodine	XXXXXX	XXXXXX	XXXXXX	XXXXXX
Cerium	XXXXXX	XXXXXX	XXXXXX	XXXXXX
Chromium	XXXXXX	XXXXXX	XXXXXX	XXXXXX
Strontium	XXXXXX	XXXXXX	XXXXXX	XXXXXX
Scandium	XXXXXX	XXXXXX	XXXXXX	XXXXXX

The centroid is used by the computer to identify the specific isotope with which it is associated.

Total CPM refers to all counts from a specific isotope in the region of interest, accumulated during the counting time--in this case one minute.

APPENDIX

REVISED SA

CORRECTED MICROSPHERE COUNTS
 NUMBER OF ISOTOPES ASSUMED TO BE 5
 ISOTOPES 1125 CE145 CR141 SR85 SC46
 ENTER SUBJECT ID F-30
 ENTER EXPERIMENT DATE 27 JUNE 78
 ENTER ASSAY DATE 30 JUNE 78
 ISOTOPE 1 1125
 ENTER VOLUME COUNTED .003
 ENTER VOLUME INJECTED .13
 ENTER # SAMPLE IDS 2
 ENTER SAMPLE IDS 1 2
 ISOTOPE 2 CE145
 ENTER VOLUME COUNTED .003
 ENTER VOLUME INJECTED .10
 ENTER # SAMPLE IDS 2
 ENTER SAMPLE IDS 3 4
 ISOTOPE 3 CR141
 ENTER VOLUME COUNTED .003
 ENTER VOLUME INJECTED .28
 ENTER # SAMPLE IDS 2
 ENTER SAMPLE IDS 5 6
 ISOTOPE 4 SR85
 ENTER VOLUME COUNTED .003
 ENTER VOLUME INJECTED .25
 ENTER # SAMPLE IDS 2
 ENTER SAMPLE IDS 7 8
 ISOTOPE 5 SC46
 ENTER VOLUME COUNTED .003
 ENTER VOLUME INJECTED .60
 ENTER # SAMPLE IDS 2
 ENTER SAMPLE IDS 9 10
 ENTER # REFERENCE SAMPLES 14
 ENTER SAMPLE IDS FOR REFERENCE 50 -63
 ENTER WITHDRAWAL RATE, REFERENCE SAMPLE 15.3
 ENTER # SAMPLES FROM RESIDUE 11
 ENTER SAMPLE IDS 11 -15 150 -155
 ENTER # SAMPLES COUNTED PREVIOUS TO ASSAY
 ENTER # SAMPLES AND # DAYS ELAPSED 77 3
 ENTER SAMPLE IDS 1 -10 11 -15 150 -155 50 -63 101 -139 141 -143
 ENTER # SAMPLES AND # DAYS ELAPSED 0 0
 ENTER # SAMPLES IN GROUP 1 0
 ENTER SAMPLE IDS 271 -273
 ENTER # SAMPLES IN GROUP 2 0
 REF CPM 147707 194252
 CORR CPM 15794. 94106.
 SYPINGE .28140E+07 .35366E+07 .32361E+07 .41069E+07 .69244E+07

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APPENDIX C
SAMPLE TAPE 2

Counter Location		Wt. (grams)		Io-125		Ce-141		Cr-51		Sr-85		Sc-46	
Code	ID	ID		Assay	date	Exp.	date	Assay	date	Exp.	date	Assay	date
S	252	52	2.9500			3939.5		7080.8		3671.5		3705.4	
						1.2937		.39024		.23733		.12607	
S	253	53	6.2790			8699.7		18804.		10315.		9971.9	
						1.3422		.38332		.31326		.15940	
S	254	54	7.2590			7785.1		11460.		5652.8		7613.5	
						1.0389		.55668		.18650		.10527	
S	255	55	6.3860			7444.8		12441.		5662.0		5113.6	
						1.0324		.28953		.15455		.73488E-01	
S	256	56	5.0220			6176.7		10083.		6252.5		6974.2	
						1.1915		.32641		.23742		.13938	
S	257	57	6.3780			9501.1		15414.		7733.1		6549.8	
						1.3392		.36435		.21440		.95578E-01	
S	258	58	1.0710			1723.2		2277.9		3175.2		3274.9	
						1.5587		.34560		.56535		.30690	
S	259	59	.93400			1289.3		2848.4		3558.0		5198.3	
						1.3373		.49582		.72644		.55861	
S	260	60	.50400			803.88		2074.8		2776.3		2908.1	
						1.5451		.66930		1.0504		.99506	
S	261	61	2.3790			1753.9		2830.6		1446.7		1138.9	
						.71418		.19345		.11596		.48049E-01	
S	262	62	2.1480			3026.0		5820.2		6280.4		6615.9	
						1.3647		.44053		.55756		.30914	
S	263	63	.66700			848.91		991.22		1101.5		1884.0	
						1.2329		.24161		.33779		.28350	
S	264	64	2.6880			2560.2		4315.7		3289.3		3326.6	
						.92267		.26103		.23193		.12421	
S	265	65	7.2590			10252.		23342.		27377.		26105.	
						1.3682		.52294		.71918		.38860	
S	266	66	14.698			17780.		29916.		19025.		18804.	
						1.1725		.33109		.24696		.12848	
C	1	300	86.700			.10920E+06		.28742E+06		.14824E+06		.14835E+06	
						1.2202		.38896		.32605		.17174	
S	F-30	27	JUNE 7030	JUNE 70	1900.5	520.92		577.50		302.17		201.3	
Subject	Assay	date	Exp.	date	Assay	date	Exp.	date	Assay	date	Exp.	date	Assay

Code: S= individual assay
 G= group assay
 F= cardiac output data

REFERENCES

1. Domenach, R.J., J.I.E. Hoffman, M.I.M. Noble, K.B. Saunders, J.R. Henson, and S. Subijanto, "Total and regional coronary blood flow measured by radioactive microspheres in conscious and anesthetized dogs," Circulation Research 25:581-596, 1969.
2. Greenlees, K. J., C.M. Oloff, W.J. Buehring, and K.C. Smith, "A technique for the injection of radioactive tracer microspheres during acceleration stress," (in press), Aviation, Space, and Environmental Medicine. (AMRL-TR-78-114)
3. Heymann, M.A., B.D. Payne, J. Hoffman, and A.M. Rudolph, "Blood flow measurement with radio-nuclide labelled particles," 1978 (in press).
4. Lederer, C.M., J.M. Hollander, and I. Perlman, Table of Isotopes, 6th ed., John Wiley and Sons Inc., New York, 1967.
5. Oloff, C.M., and W.L. Finch, Subhuman Primate Restraint System, AMRL-TR-78-88, Aerospace Medical Research Laboratory, Wright-Patterson AFB, Ohio, December 1978.
6. Rudolph, A.M., and M.A. Heymann, "The circulation of the fetus in utero: Methods for studying distribution of blood flow, cardiac output, and organ blood flow." Circulation Research 21:163-184, 1967.
7. Sostre, S., J.A. Kennealy, J.S. Kirkland, C.M. Oloff, A.A. Karl, and M.A. Franey, "Cerebral blood flow in baboons under positive acceleration." Aerospace Medical Assn., Annual Sci. Mtgs., 75-76, 1977.